Honulactones: New Bishomoscalarane Sesterterpenes from the Indonesian Sponge *Strepsichordaia aliena*

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From the dichloromethane/2-propanol (1:1) extract of the Indonesian marine sponge *Strepsichordaia aliena*, twelve new 20,24-bishomoscalarane sesterterpenes, honulactones A–L (**1–12**) were isolated. Molecular structures were secured by spectroscopic methods, accurate mass measurements, and X-ray analysis. Honulactones A (**1**), B (**2**), C (**3**), and D (**4**) exhibit cytotoxycity against P-388, A-549, HT-29, and MEL-28 (IC₅₀ 1 μ g/mL).

Sponges of the order Dictyoceratida are prominent members of the Indo-Pacific coral reefs and often good sources of scalarane-based sesterterpenes.^{1–5} Some sesterterpenes exhibit potentially useful biological properties such as antiinflamatory,⁶ cytotoxic,^{3,5,7} antifeedant,⁸ platelet aggregation,⁹ and ichthyotoxic.¹⁰ Some scalarane sestertepenoids include alkylated derivatives which can be further divided into four known skeletal types.¹¹

Homoscalaranes are methylated at C-20 or C-24, while methylation at C-20 and C-24 characterizes bishomoscalaranes.¹² We now report isolation and structural

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elucidation of eleven new bishomoscalaranes, honulactones A–L (1-12),¹³ from the Indonesian sponge *Strepsichordaia aliena*.¹⁴ Honulactones A (1), B (2), and E–H (5–8) represent a new skeletal system possessing a cyclopropane ring. Compounds 1 and 2 differ in the orientation of the CH₃-26 group as do 5 and 6 as well as 7 and 8. Furthermore, 5 and 6 are pentanoates rather than butanoate esters of the C-12 hydroxyl. Finally, compounds 7 and 8 are the corresponding C-16 hydroxylated analogues of honulactones A and B.

(13) Honu is the Hawaiian word for turtle, which reflects on the collection site at Turtle Bay, eastern Indonesia. Isolation procedure: The freeze-dried sponge (81.0 g) was extracted in DCM:IPA (1:1; 1.0 L) overnight, filtered, and concentrated under reduced pressure until dryness to yield 3.03 g of crude extract. The crude extract was loaded atop a Sephadex LH-20 column (30 \times 2.5 cm) equilibrated in dichloromethane. The column was eluted using a gradient profile as follows: (1) dichloromethane (DCM), DCM:acetone (1:1), and methanol. Eight major fractions (A-H) were collected and concentrated to dryness. Reverse-phase HPLC (Phenomenex Ultracarb 10 ODS 30; 250 \times 22 mm; 80% aq MeCN to 100% MeCN in 40 min at 6.0 mL/min and monitoring at 220 nm) of fraction B afforded six fractions [fr 1 (35.2 mg), fr 2 (77.2 mg), fr 3 (32.4 mg), fr 4 (118.1 mg), fr 5 (44.7 mg), fr 6 (90.3)]. Fractions 3, 4, 5, and 6 were further separated by normal-phase HPLC (Microsorb Si; 300×7.0 mm. Solvent A = hexanes; solvent B = 1:1 hexanes/2-propanol. Starting with solvent A at 0 min to 100% solvent B in 35 min at 2.0 mL/min and monitoring at 220 nm) to yield honulactone A (7.1 mg), honulactone B (6.0 mg), honulactone Č (2.8 mg), honulactone D (2.0 mg), honulactone E (4.5 mg), honulactone F (3.4 mg), honulactone I (2.4 mg), honulactone J (2.6 mg), honulactone K (2.3 mg), and honulactone L (1.6 mg). Reverse-phase HPLC (Phenomenex Ultracarb 10 ODS 30; 250 \times 22 mm; 80% aq MeCN to 100% MeCN in 40 min at 6.0 mL/min and monitoring at 220 nm) of fraction C afforded nine fractions. Fraction 4 was further separated by normalphase HPLC (Microsorb Si; 300 \times 7.0 mm. Solvent A = hexanes; solvent B = 1:1 hexanes/2-propanol. Starting with solvent A at 0 min to 100% solvent B in 35 min at 2.0 mL/min and monitoring at 220

(14) The sponge was collected at Turtle Bay, Sangakali, eastern Indonesia, at a depth of 23 m, in March 1996 (2° 04′ 59° N, 118° 23′ 41° E). In life, the sponge is fan-shaped to palmitate-digitate, with 2 mm diameter oscules on one surface; the opposite surface has radiating channels, and both surfaces are covered with small conules. The texture is quite tough, but very flexible, the external color in life, maroonpurple, interior cream. The skeleton consists of simple radiating cored primary fibers and golden vermiform tertiary fibers which are linked by short junctions. The surface has a layer of sand-grains on it. The sponge is closely comparable to *Strepsichordaia aliena* (order Dictyoceratida, Family Thorectidae, Subfamily Phyllosponginae). A voucher specimen has been deposited in the Natural History Museum, London (BMNH 1999.7.12.1).

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The distinctive common feature of honulactones C (3)/ D (4), I (9)/ J (10), and K (11)/L (12) is C-20 oxidation. Compounds 3 and 4 are epimers at C-24. Honulactone I (9) and J (10) are C-12 pentanoate ester. Finally, honulactones J (10) and K (11) are the C-20 propanoates rather than acetate esters as well as epimers at C-24. Compounds 3, 4, and 9–12 represent further examples of C-20 oxidized bishomoscalaranes.¹⁵

Honulactone A (1)¹⁶ was obtained as a colorless solid with a molecular formula of C₃₁H₄₆O₅ as established by HRFABMS, m/z [M + H]⁺ 499.3431. The ¹H NMR spectrum of 1 indicated six methyl groups: three methyl singlets at δ 0.79, 0.88, 1.17; and three methyl doublets at δ 1.08, 1.18, and 1.36. ¹H⁻¹H COSY and 1D TOCSY experiments revealed that CH_3 -27 (d, J = 6 Hz) resonating at δ 1.08 was coupled to CH-20 resonating at δ 0.7 (ddq, J = 4, 6.4, 8.4 Hz), and the latter was coupled to two cyclopropane protons resonating at δ 0.58 and -0.49. In addition, CH₃-4' absorbing at δ 1.18 (d, J = 6.9 Hz) was coupled to CH-3' at δ 4.10 and CH₃-26 resonating at δ 1.36 (d, J = 6.8 Hz) was coupled to CH-24 at δ 4.78.



The IR and ¹³C spectra indicated the presence of an α,β -unsaturated γ -lactone (v_{max} 1742 cm⁻¹; δ_{C} 171.3, 164.1, and 132.7) and a hydroxy ester (v_{max} 1671 cm⁻¹; $\delta_{\rm C}$ 64.2, 171.5). The ¹³C NMR spectrum showed four quaternary carbons [C-13 (8 38.4), C-8(8 37.8), C-10 $(\delta 37.2)$, C-4 $(\delta 22.6)$] and three tertiary methyl groups (δ 13.9, 16.8, 21.4 at C-22, C-21, and C-23, respectively). These chemical shifts were suggestive of axial methyl groups at the ring junctions C-8, C-10, and C-13 in an all-trans A-B-C-D ring system in accordance with wellknown assignments of other scalarane sesterterpenes¹⁷ and triterpenes.18

The proton signals at $\delta_{\rm H}$ 0.58, -0.49, 4.10, 4.78, and 5.61 were key elements in the structure elucidation. HMBC correlation between H-3' (δ 4.10) and C-2' (δ 43.4)/ C-4' (δ 22.25) further confirmed the existence of a 3-hydroxybutanoate moiety attached to the carbon-bearing oxygen at C-12 [HMBC correlation between H-12 (δ 5.61) and C-1' (δ 171.5)] on ring E. The γ -lactone unit was evidently fused to ring D as seen by HMBC correlations between H-12 to C-18 (δ 132.7), H-16 (δ 2.21, 2.37) to C-17 (δ 164.1)/C-18, and H-15 (δ 1.54, 1.91) to C-17. Further evidence of the γ -lactone on ring D was obtained from HMBC correlations between H-24 (δ 4.78) to C-17/ C-18 and H-26 (δ 1.36) to C-17. The cyclopropane signals at C-19 ($\delta_{\rm H}$ 0.58, -0.49; $\delta_{\rm C}$ 13.6) were correlated to C-20 ($\delta_{\rm H}$ 0.70; $\delta_{\rm C}$ 13.4), and both were connected to ring A through C-4 ($\delta_{\rm C}$ 22.6): H-19_{cis} (δ 0.58) and H-19_{trans} (δ -0.49) showed correlations to C-3/C-4/C-5; and H-20 (δ 0.7) showed correlations to C-4, C-5, and C-19. Finally, the C-27 methyl doublet (δ 1.08) at C-20 on the cyclopropane ring was secured based on HMBC correlations between H-27 to C-4, C-19, and C-20.

The relative configuration of **1** was deduced from its NOESY spectrum. The small J-value observed for H-12 indicated an equatorial hydrogen. The CH₃-26 group was assigned β -orientation on the basis of a strong NOE observed between H-16eq and H-26. Also, 1D-NOE experiments provided further evidence for β -orientation: irradiation of CH₃-26 produced a positive NOE on H-24 and H-16_{eq}. The all-trans A-B-C-D ring system was also confirmed by cross-peaks in the NOESY spectrum: $H-11_{ax}$ to CH_3-23_{ax} and $H-15_{ax}$ to CH_3-23_{ax} . The C-20 cyclopropane methine carbon was assigned β -orientation, since a strong NOESY cross-peak was observed between H-20 to H-22. The relative configuration of H-20 group was assigned as $20S^*$: irradiation of H-27 produced a positive NOE on H-3_{eq}, H-20, H-22, and H-19_{trans}.

¹H, ¹³C, ¹H-¹H COSY, 1D-TOCSY, and HMBC NMR spectra of honulactones B (2),¹⁹ E (5),,²⁰ F (6),²¹ G (7),²² and H (8)²³ display the same H–H and C–C sequences seen in 1: an α,β -unsaturated γ -lactone, an all-trans

(16) Honulactone A (1): Colorless crystalline solid, 7.1 mg (0.0088% based on dry weight); $[a]_D = +73.2^{\circ}$ (*c* 0.71, CH₂Cl₂). HRFABMS *m*/*z* 499.3431 [M + H]⁺ (C₃₁H₄₇O₅, Δ -1.5 ppm). IR (thin film) v_{max} 3448, 2930, 1742, 1671, 1383, 1324, 1288, 1176, 1064 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ 1.70 (dt, *J* = 3, 13 Hz, H-1_{eq}), 0.71 (m, H-1_{ax}), 1.44 (m, H₂-2), 1.5 (m, H-3_{eq}), 1.23 (m, H-3_{ax}), 1.37 (m, H-5_{ax}), 1.05 (m, H₂-6), 1.77 (dt, *J* = 3, 12 Hz, H-7_{eq}), 0.97 (m, H-7_{ax}), 1.19 (dd, *J* = 2, 14 Hz, H-9_{ax}), 2.09 (dt, *J* = 3, 15 Hz, H-11_{eq}), 1.7 (ddd, *J* = 2, 15, 15 Hz, H-11_{eq}), 1.52 (m, H-14_{ax}), 1.191 (dd, *J* = 6, 13 Hz, H-15_{eq}), 1.54 (m, H-15_{ax}), 2.37 (m, H-6_{eq}), 2.21 (m, H-16_{ax}), 0.58 (dd, *J* = 4, 6, 8.4 Hz; H-20), 0.88 (s, H₃-21), 0.79 (s, H₃-22), 1.17 (s, H₃-23), 4.78 (q, *J* = 6.6 Hz, H-24), 1.36 (d, *J* = 6.8 Hz, H₃-26), 1.08 (d, *J* = 6.4 Hz, H₃-27), 2.37 (m, H-2'a), 2.31 (m, H-2'b), 4.10 (m, H-3), 3.06 (d, *J* = 3, 11.4 k, Hc-3), and 1.18 (d, *J* = 6.9 Hz, H₃-4). ¹³C NMR (100 MHz, CDCl₃) δ 39.7 (C-1), 21.1 (C-2), 33.0 (C-3), 22.6 (C-4), 50.2 (C-5), 17.5 (C-6), 40.0 (C-7), 37.8 (C-8), 51.4 (C-9), 37.2 (C-10), 21.1 (C-11), 74.5 (C-12), 38.4 (C-13), 51.2 (C-14), 16.8 (C-15), 24.0 (C-16), 164.1 (C-17), 132.7 (C-18), 13.6 (C-19), 13.4 (C-20), 16.8 (C-21), 13.9 (C-22), 21.4 (C-22), 77.9 (C-24), 171.3 (C-25), 18.6 (C-26), 13.1 (C-27), 171.5 (C-1), 43.4 (C-2), 64.2 (C-3), and 22.3 (C-4'). (17) Crews, P.; Naylor, S. *Fortschr. Chem. Org. Naturst.* **1985**, 48, 203–269 (16) Honulactone A (1): Colorless crystalline solid, 7.1 mg (0.0088%

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(19) Honulactone B (2): Colorless crystalline solid, 6.0 mg (0.0074% based on dry weight); $[\alpha]_D = +77^\circ$ (c 0.6, CH₂Cl₂). HRFABMS m/z499.3417 $[M + H]^+$ (C₃₁H₄₇O₅, Δ 1.3 ppm). IR (thin film) v_{max} 3448, 2962, 2930, 2869, 1742, 1671, 1458, 1384, 1268, 1175, 1021 cm $^{-1}$. $^1\mathrm{H}$ NMR (400 MHz, CDCl₃) Proton chemical shifts for **2** are within ± 0.03 ppm of the values for **1**. ¹³C NMR (100 MHz, CDCl₃) Carbon chemical shifts for **2** are identical to **1** except for δ 51.5 (C-14), 16.6 (C-15), 24.3 (C-16), and 78.1 (C-24).

(20) Honulactone E (5): Colorless crystalline solid, 4.5 mg (0.0056% based on dry weight); $[a]_D = +105.2^{\circ}$ (*c* 1.5, CH₂Cl₂). HRMS (DCI) *m/z* 530.383397 [M + NH₄]⁺ (C₃₂H₅₂NO₅, Δ 2.2 ppm). IR (thin film) v_{max} 3500, 2900, 1740, 1680 cm⁻¹. ¹H NMR (500 MHz, CDCl₃) δ 1.70 (m, H-1_{eq}), 0.71 (m, H-1_{ax}), 1.45 (m, H₂-2), 1.53 (m, H-3_{eq}), 1.22 (m, H-3_{ax}), 1.37 (m, H-5_{ax}), 1.03 (m, H₂-6), 1.76 (dt, J = 3, 12 Hz, H-7_{eq}), 0.97 (m, 1.37 (m, H-5_{ax}), 1.03 (m, H₂-0), 1.76 (dt, J = 3, 12 H2, H-7_{eq}), 0.97 (m, H-7_{ax}), 1.19 (m, H-9_{ax}), 2.10 (dt, J = 3, 15 Hz, H-11_{eq}), 1.71 (m, H-11_{ax}), 5.60 (br t, J = 2.7 Hz, H-12_{eq}), 1.52 (m, H-14_{ax}), 1.90 (dd, J = 7, 13 Hz, H-15_{eq}), 1.55 (m, H-15_{ax}), 2.37 (m, H-16_{eq}), 2.22 (m, H-16_{ax}), 0.57 (dd, J = 4.2, 8.7 Hz, H-19_{cts}), -0.49 (t, J = 5.2 Hz, H-19_{trans}), 0.69 (m, H-20), 0.87 (s, H₃-21), 0.78 (s, H₃-22), 1.17 (s, H₃-23), 4.77 (q, J = 6.8 Hz, H-24), 1.36 (d, J = 6.8 Hz, H₃-26), 1.07 (d, J = 6.3 Hz, H₃-27), 2.38 (m, H24), 0.96 (m, H24), 0.96 (m, H26), 0.97 (m, 0.97), 0.97 (13), 51.1 (C-14), 16.8 (C-15), 24.0 (C-16), 164.1 (C-17), 132.7 (C-18), 13.5 (C-19), 13.3 (C-20), 16.8 (C-21), 13.7 (C-22), 21.4 (C-23), 77.9 (C-24), 171.3 (C-25), 18.6 (C-26), 13.1 (C-27), 171.1 (C-1'), 41.5 (C-2'), 69.4 (C-3'), 29.3 (C-4'), and 10.0 (C-5').

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Figure 1. ORTEP drawing of honulactone B (2) and D (4).

A–B–C–D ring system, and a methylcyclopropane. However, there were small variations on the structural motif: (1) in compound **2**, the CH₃-26 group was α -oriented; (2) honulactones E (**5**) and F (**6**) were the 3-hydroxypentanoate ester homologues of honulactone A and B having a 26 β -CH₃ in compound **5**, while a 26 α -CH₃ in **6**; (3) honulactones G (**7**) and H (**8**) were the corresponding 16 α -OH analogues of **1** and **2** possessing a 26 β -CH₃ in compound **7**, while a 26 α -CH₃ in **8**. The relative configuration of all compounds was secured by 1D-NOE experiments. The relative configuration and gross struc-

(21) Honulactone F (**6**): Colorless crystalline solid, 3.4 mg (0.0042% based on dry weight); $[\alpha]_D = +81.5^{\circ}$ (*c* 1.1, CH₂Cl₂). HRMS (DCI) *m/z* 530.382715 [M + NH₄]⁺ (C₃₂H₅₂NO₅, Δ 3.5 ppm). IR (thin film) v_{max} 3450, 2890, 1730, 1650 cm⁻¹. ¹H NMR (500 MHz, CDCl₃) Proton chemical shifts for **6** are within ±0.02 ppm of the values for **5**. ¹³C NMR (125 MHz, CDCl₃) Carbon chemical shift values for **6** are identical to **5** except for δ 51.4 (C-14), 16.5 (C-15), 24.3 (C-16), and 78.1 (C-24).

(22) Honulactone G (7): Colorless crystalline solid, 1.7 mg (0.0021% based on dry weight); $[a]_D = +85.7^{\circ}$ (c 0.85, CH₂Cl₂). HRMS (DC1) m/z 532.366487 [M + NH₄]⁺ (C₃₁H₅₀NO₆, Δ -5.0 ppm). IR (thin film) v_{max} 3500, 3400, 2990, 1735, 1660 cm^{-1.} ¹H NMR (500 MHz, CDCl₃) δ 1.70 (m, H-1_{eq}), 0.70 (m, H-1_{ax}), 1.50 (m, H-2_{eq}), 1.45 (m, H-2_{ax}), 1.50 (m, H-3_{eq}), 1.25 (m, H-3_{ax}), 1.41 (m, H-5_{ax}), 1.05 (m, H₂-6), 1.70 (m, H-1_{eq}), 0.70 (m, H-1_{ax}), 2.11 (dt, J = 3, 15 Hz, H-11_{eq}), 1.70 (m, H-1_{ax}), 5.62 (br t, J = 2.7 Hz, H-12_{eq}), 1.80 (m, H-14_{ax}), 1.85 (m, H-2₁₅), -0.49 (t, J = 5.2 Hz, H-19_{trans}), 0.68 (m, H-20), 0.86 (s, H₃-21), 0.79 (s, H₃-22), 1.14 (s, H₃-23), 5.08 (q, J = 6.8 Hz, H-24), 1.41 (d, J = 6.8 Hz, H₃-26), 1.08 (d, J = 6.3 Hz, H₃-27), 2.34 (m, H₂-2), 4.10 (m, H-3), 3.03 (s, HO-3'), and 1.20 (d, J = 6.9 Hz, H₃-4'). ¹³C NMR (125 MHz, CDCl₃) δ 39.7 (C-1), 21.2 (C-2), 33.0 (C-3), 22.6 (C-4), 50.2 (C-5), 17.4 (C-6), 39.9 (C-7), 36.7 (C-8), 51.4 (C-9), 37.8 (C-10), 21.1 (C-11), 74.2 (C-12), 39.0 (C-13), 45.9 (C-14), 27.7 (C-15), 61.5 (C-16), 162.0 (C-17), 135.5 (C-18), 13.6 (C-3), 13.0 (C-26), 13.1 (C-27), 171.5 (C-1'), 43.4 (C-2'), 64.3 (C-3'), and 22.4 (C-4').

(23) Honulactone H (8): Colorless crystalline solid, 1.5 mg (0.0019% based on dry weight); $[\alpha]_D = +78.3^{\circ}$ (*c* 0.75, CH₂Cl₂). HRMS (DC1) *m/z* 532.364078 [M + NH₄]⁺ (C₃₁H₅₀NO₆, Δ -0.5 ppm). IR (thin film) v_{max} 3505, 3200, 2900, 1745, 1670 cm⁻¹. ¹H NMR (500 MHz, CDCl₃) Proton chemical shifts for 8 are within ±0.03 ppm of the values for 7 except for δ 4.90 (q, J = 6.8 Hz, H-24), 1.52 (d, J = 6.8 Hz, H₃-26), 2.37 (m, H-2'a), and 2.36 (m, H-2'b). ¹³C NMR (125 MHz, CDCl₃) Carbon chemical shift values for 8 are identical to 7 except for δ 62.9 (C-16), 160.8 (C-17), 78.9 (C-24), and 19.8 (C-26).

(24) Suitable crystals of honulactone B (2) for X-ray analysis were obtained from isooctane/dichloromethane. The compound crystallized in the tetragonal space group $P4_32_12$ with a unit cell having the dimensions a = 29.924 (1) Å, b = 29.924 (1) Å, c = 7.3309 (4) Å, and a calculated density of 1.009 g cm⁻³. A colorless crystal (0.40 × 0.10 × 0.10 mm³) mounted on a thin glass rod was used for the data collection. A total of 1321 frames of data were taken on a BRUKER SMART CCD Area Detector System equipped with a 3 kW sealed tube (Mo $\kappa\alpha$) X-ray generator. A narrow-frame method was used with a scan widths of 0.3° in ω and an exposure time of 30 s/frame. Frames were integrated to yield a total of 18626 reflections of which 2577 were independent ($R_{\rm int} = 7.31\%$), and 2343 were above $4\sigma(F)$. The structure was solved by direct methods and refined by full-matrix least squares on F^2 using anisotropic displacement parameters for all non-hydrogen atoms. At final convergence, $R_1 = 8.63\%$ and GOF = 1.040 for 340 parameters. Additional X-ray data are available in the Supporting Information and the Cambridge Crystallographic Data file.



ture of honulactone B (**2**) was also secured by X-ray analysis.²⁴ An ORTEP drawing is shown in Figure 1.

Initial inspection of the ¹H NMR spectrum of honulactone C (**3**)²⁵ indicated the absence of cyclopropane resonances (δ 0.7, 0.50, and -0.49) and the appearance of new signals at δ 2.03 and 5.34 attributed to a CH₃CO and CH–OR units. The latter functional units were also confirmed by the molecular formula of C₃₃H₅₀O₇ as established by HRFABMS, *m*/*z* 559.3615. IR absorption at 1738 cm⁻¹ indicated an α , β -unsaturated γ -lactone, and further evidence of this functional group was obtained from the ¹³C NMR spectrum (δ 171.3, 164.3, 132.6). Additional IR absorptions at 3498 and 1690 cm⁻¹ were also indicative of a hydroxyl and acetate groups: $\delta_{\rm H}$ 3.07 for 3'-OH; $\delta_{\rm H}$ 4.07, $\delta_{\rm C}$ 64.2 for the carbinol methine at C-3'; and $\delta_{\rm H}$ 2.02, $\delta_{\rm C}$ 21.8 for the methyl ketone (HMBC correlation between $\delta_{\rm H-29}$ 2.02 to $\delta_{\rm C-28}$ 170.3).



The ¹³C NMR spectrum showed four quaternary carbons and four tertiary methyl groups with a gem-methyl/ ethyl group at C-4 and axial methyl groups at the ring junctions C-8, C-10, and C-13.^{10,15,26} Furthermore, signals at $\delta_{\rm C}$ 17.7, 19.9, 21.0 could be attributed to carbons C-2, C-6, and C-11 located γ to axial methyl groups, while the carbon signals at δ 40.3 and 42.3 (δ_{H-1} 0.62, 1.64; δ_{H-7} 0.90, 1.81) can be assigned to C-1 and C-7 located β to the axial methyl groups. The relative configuration of 3 was deduced from the NOESY spectrum. The J-value of H-12 indicated that it was an equatorially oriented, which was also confirmed by NOESY cross-peak between H-12 and H-11_{ax}/H-11_{eq}/H-23. The relative configuration of CH₃-26 group was assigned β -orientation based on a strong NOE observed between $H-16_{eq}$ and H-26. The all-trans A-B-C-D ring system was also confirmed by crosspeaks observed in the NOESY spectrum: H-9_{ax} to H-1_{ax},

H-5_{ax}, H-14_{ax}; H-11_{ax} to CH₃-21, CH₃-22, CH₃-23; and H-15_{ax} to CH₃-21, CH₃-23. The substituted ethyl sidechain at C-4 has β -orientation, since a strong NOESY cross-peak was observed between H-20 to CH₃-22, and CH₃-19_{ax} to H-6_{eq}/H-3_{eq}. The relative configuration of the C-20 acetoxy group was assigned as 20*R** on the basis of a cross-peak between the CH₃CO and H-6_{eq}/H-6_{ax}, and CH₃-27 and H-3_{ax}.

Spectral data (¹H,¹³C, COSY, 1D-TOCSY, HMQC, and HMBC) identified honulactones D (**4**)²⁷ as the C-26 epimer (26 α -Me) of **3**, honulactone I (**9**)²⁸ and J (**10**)²⁹ as the 3-hydroxypentanoate ester (26 β -Me and 26 α -Me, respectively) homologues of honulactone C, and honulactones J (**11**)³⁰ and K (**12**)³¹ as the C20-propionate ester

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(27) Honulactone D (**4**): Colorless crystalline solid, 4.5 mg (0.0056% based on dry weight); $[\alpha]_D = +62.0^{\circ}$ (*c* 0.25, CH₂Cl₂). HRFABMS *m/z* 559.3618 [M + H]⁺ (C₃₃H₅₁O₇, Δ 3.0 ppm). IR (thin film) v_{max} 3498, 2969, 1738, 1732, 1672, 1254, 1033 cm⁻¹. ¹H NMR (500 MHz, CDCl₃) Proton chemical shifts for **4** are within ±0.05 ppm of the values for **3** except for δ 2.33 (d, J = 6 Hz, H₂-2'). ¹³C NMR (125 MHz, CDCl₃) (C-24).

(28) Honulactone I (9): Colorless crystalline solid, 2.4 mg (0.003% based on dry weight); $[\alpha]_D = +83.4^{\circ}$ (c 0.96, CH₂Cl₂). HRMS (FAB) m/z 573.37913 [M + H]⁺ ($C_{34}H_{53}O_7, \Delta - 4.5$ ppm). IR (thin film) v_{max} 3490, 2970, 1725, 1660, 1360, 1025 cm⁻¹. ¹H NMR (500 MHz, CDCl₃) δ 1.72 (m, H-1_{eq}), 0.64 (ddd, J = 4, 13, 13 Hz, H-1_{ax}), 1.45 (m, H-2_{eq}), 1.35 (m, H-2_{ax}), 1.66 (m, H-3_{eq}), 1.00 (ddd, J = 4, 14, 14 Hz, H-3_{ax}), 0.96 (m, H-5_{ax}), 1.75 (m, H-6_{eq}), 1.43 (m, H-6_{ax}), 1.81 (dt, J = 3, 13 Hz, H-7_{eq}), 0.91 (m, H-7_{ax}), 1.14 (dd, J = 3, 13 Hz, H-9_{ax}), 2.04 (dt, J = 3, 13 Hz, H-11_{eq}), 1.69 (m, H-11_{ax}), 5.60 (br t, J = 2.9 Hz, H-12_{eq}), 1.50 (m, H-14_{ax}), 1.89 (dt, J = 7, 13 Hz, H-15_{eq}), 1.55 (m, H-15_{ax}), 2.36 (d, J = 15 Hz, H-16_{eq}), 2.19 (m, H-16_{ax}), 0.96 (s, H₃-19), 5.35 (q, J = 6.9 Hz, H-20), 0.86 (s, H₃-21), 0.86 (s, H₃-22), 1.17 (s, H₃-23), 4.76 (q, J = 6.9 Hz, H-24), 1.35 (d, J = 7.0 Hz, H₃-26), 1.08 (d, J = 7.0 Hz, H₃-27), 2.03 (s, CH₃CO), 2.37 (dd, J = 3, 16 Hz, H-2'a), 2.29 (dd, J = 9, 16 Hz, H-2'b), 3.82 (m, H-3), 2.95 (s, HO-3'), 1.47 (m, H₂-4'), and 0.93 (t, J = 7.6 Hz, H₃-5). ¹³C NMR (125 MHz, CDCl₃) δ 40.3 (C-1), 17.8 (C-2), 39.0 (C-3), 39.2 (C-4), 58.9 (C-5), 20.0 (C-6), 42.4 (C-7), 37.5 (C-8), 53.8 (C-9), 37.1 (C-10), 21.0 (C-11), 74.5 (C-12), 38.3 (C-13), 51.0 (C-14), 16.8 (C-15), 24.0 (C-16), 164.0 (C-17), 132.7 (C-18), 23.2 (C-19), 73.1 (C-20), 16.6 (C-21), 16.5 (C-22), 21.4 (C-23), 77.8 (C-24), 171.2 (C-25), 18.6 (C-2), 69.4 (C-3'), 29.3 (C-4'), and 10.0 (C-5'). (29) Honulactone J (10): Colorless crystalline solid, 2.6 mg (0.0032%)

(29) Honulactone J (**10**): Colorless crystalline solid, 2.6 mg (0.0032% based on dry weight); $[\alpha]_D = +80.7^\circ$ (c 0.67, CH₂Cl₂). HRMS (DCI) m/z 590.403408 [M + NH₄]⁺ (C₃₄H₅₆NO₇, Δ 3.8 ppm). IR (thin film) v_{max} 3500, 2990, 1750, 1650 cm⁻¹. ¹H NMR (500 MHz, CDCl₃) Proton chemical shifts for **10** are within ±0.05 ppm of the values for **9** except for δ 2.36 (m, H-16_{eq}), and 2.25 (m, H-16_{ax}). ¹³C NMR (125 MHz, CDCl₃) Carbon chemical shifts for **10** are identical to **9** except for δ 78.1 (C-24).

homologues of **3**/**4** having a 26β -CH₃ in compound **11**, while a 26α -CH₃ in **12**. The relative configuration of all compounds was determined by 1D-NOE experiments. Additional support for the relative configuration and gross structure of honulactone D (**4**) was secured by X-ray analysis.³² An ORTEP drawing is shown in Figure 1.

Evaluation of honulactones A-D (1–4) against P-388 (ATCC: CCL 46), A-549 (ATCC: CCL 8), HT-29 (ATCC: HTB 38), and MEL-28 (ATCC: HTB 72) showed IC₅₀ values of 1 µg/mL for all compounds. No cytotoxic evaluation was performed on compounds **5–12**. The cancer-cell growth-inhibitory activity shown by sesterterpenes similar to compounds **1–4** is likely the result of Michael-type additions of biosynthetic thiol and/or related groups to the α,β -unsaturated γ -lactone system.^{7e,33}

Cytotoxicity Testing. Cytotoxicity assays were carried out by Instituto Biomar, S. A., Madrid, Spain.

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Supporting Information Available: ¹H and ¹³C NMR spectra for all compounds, and X-ray data and ORTEP projections for compounds **2** and **4**. This material is available free of charge via the Internet at http://pubs.acs.org.

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(30) Honulactone K (11): Colorless crystalline solid, 2.3 mg (0.0028% based on dry weight); $[\alpha]_D = +90.1^{\circ}$ (c 0.77, CH₂Cl₂). HRMS (DCI) m/z 590.402819 [M + NH₄]⁺ (C₃₄H₅₆NO₇, Δ 4.8 ppm). IR (thin film) v_{max} 3560, 3020, 2900, 1760, 1670 cm⁻¹. ¹ H NMR (500 MHz, CDCl₃) δ 1.66 (m, H-1_{eq}), 0.63 (ddd, J = 4, 13, 13 Hz, H-1_{ax}), 1.43 (m, H-2_{eq}), 1.33 (m, H-2_{ax}), 1.64 (m, H-3_{eq}), 1.00 (ddd, J = 4, 14, 14 Hz, H-3_{ax}), 0.95 (m, H-5_{ax}), 1.70 (m, H-6_{eq}), 1.40 (m, H-6_{ax}), 1.80 (dt, J = 3, 13 Hz, H-7_{eq}), 0.89 (m, H-7_{ax}), 1.2 (m, H-9_{ax}), 2.02 (dt, J = 3, 15 Hz, H-11_{eq}), 1.55 (m, H-11_{ax}), 5.60 (br t, J = 2.7 Hz, H-12_{eq}), 1.49 (m, H-14_{ax}), 1.89 (dd, J = 7, 13 Hz, H-15_{eq}), 1.55 (m, H-15_{ax}), 2.35 (m, H-16_{eq}), 2.19 (m, H-6_{ax}), 0.97 (s, H₃-19), 5.38 (q, J = 6.3 Hz, H-20), 0.85 (s, H₃-21), 0.86 (s, H₃-22), 1.17 (s, H₃-23), 4.77 (q, J = 6.8 Hz, H-24), 1.35 (d, J = 6.8 Hz, H-26), 1.07 (d, J = 6.3 Hz, H-24), 1.35 (d, J = 6.8 Hz, H₂, 26), 1.07 (d, J = 6.3 Hz, H-2′a), 2.30 (m, H-2′b), 4.10 (m, H-3′), 3.05 (s, HO-3′), and 1.18 (d, J = 6.3 Hz, H₃-4′). ¹³C NMR (125 MHz, CDCl₃) δ 40.3 (C-1), 17.8 (C-2), 39.0 (C-3), 39.3 (C-4), 58.9 (C-5), 20.1 (C-6), 42.3 (C-7), 37.5 (C-8), 53.8 (C-9), 37.1 (C-10), 21.0 (C-11), 74.5 (C-12), 38.3 (C-13), 51.0 (C-14), 16.7 (C-15), 24.0 (C-16), 164.1 (C-17), 132.7 (C-18), 23.2 (C-19), 72.8 (C-20), 16.5 (C-21), 16.6 (C-22), 21.4 (C-23), 77.9 (C-24), 171.3 (C-25), 18.6 (C-26), 15.8 (C-27), 173.5 (CH₃-CH₂CO), 28.5 (CH₃CH₂CO), 9.2 (CH₃CH₂CO), 171.5 (C-1'), 43.4 (C-2'), 2.9, 42.2 (C-4').

(31) Honulactone L (12): Colorless crystalline solid, 1.6 mg (0.002% based on dry weight); $[\alpha]_D = +74^{\circ}$ (c 0.8, CH₂Cl₂). HRMS (DCI) m/z 590.403456 [M + NH₄]⁺ (C₃₄H₅₆NO₇, Δ 3.8 ppm). IR (thin film) v_{max} 3565, 3025, 2910, 1770, 1660 cm⁻¹. ¹H NMR (500 MHz, CDCl₃) Proton chemical shifts for 11 are within \pm 0.05 ppm of the values for 10 except for δ 2.32 (m, H₂-2'). ¹³C NMR (125 MHz, CDCl₃) Carbon chemical shift values for 11 are identical to 10 except for δ 78.1 (C-24).

(32) Suitable crystals of honulactone D (4) for X-ray analysis were obtained from isooctane/dichloromethane. The compound crystallized in the orthorhombic space group $P2_12_12_1$ with a unit cell having the dimensions a = 7.5059(1) Å, b = 14.1990(3) Å, c = 28.7786(10) Å, and a calculated density of 1.21 g cm^-3. A colorless crystal (0.15 imes 0.10 imes0.10 mm³) mounted on a thin glass rod was used for the data collection. A total of 1321 frames of data were taken on a BRUKER SMART CCD Area Detector System equipped with a 3 kW sealed tube (Mo Ka) X-ray generator. A narrow-frame method was used with a scan widths of 0.3° in ω and an exposure time of 30 s/frame. Frames were integrated to yield a total of 9611 reflections of which 3530 were independent $(R_{int} = 11.08\%)$, and 3295 were above $4\sigma(F)$. It was impossible to cut a single crystal out of a conglomerate, so the low-resolution data were heavily compromised by reflections from small satellite crystals and discarded. The structure was solved by direct methods and refined by full-matrix least squares on F^2 using anisotropic displacement pa-rameters for all non-hydrogen atoms. At final convergence, $R_1 = 7.73\%$ and GOF = 1.042 for 361 parameters. Additional X-ray data are available in the Supporting Information and the Cambridge Crystallographic Data file.

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⁽²⁵⁾ Honulactone C (3): Colorless crystalline solid, 7.6 mg (0.0094% based on dry weight); $[\alpha]_D = +71.2^{\circ}$ (c 0.57, CH₂Cl₂). HRFABMS m/z 559.3615 [M + H]⁺ (C₃₃H₅₁O₇, Δ 3.5 ppm). IR (thin film) ν_{max} 3498, 2969, 1738, 1672, 1372, 1033 cm⁻¹. ¹H NMR (500 MHz, CDCl₃) δ 1.64 (m, H-1_{eq}), 0.62 (ddd, J = 4, 13, 13 Hz, H-1_{ax}), 1.43 (m, H-2_{eq}), 1.34 (m, H-2_{ax}), 1.63 (m, H-3_{eq}), 0.99 (ddd, J = 4, 14, 14 Hz, H-3_{ax}), 0.94 (m, H-5_{ax}), 1.74 (m, H-6_{eq}), 1.42 (m, H-6_{ax}), 1.81 (dt, J = 3, 13 Hz, H-7_{eq}), 0.90 (ddd, J = 3, 13 Hz, H-1_{eq}), 1.68 (m, H-11_{ax}), 5.58 (br t, J = 2.7 Hz, H-12_{eq}), 1.48 (d, J = 13 Hz, H-14_{ax}), 1.88 (dt, J = 3, 13 Hz, H-15_{eq}), 1.53 (m, H-15_{ax}), 2.35 (m, H-16_{eq}), 2.19 (m, H-16_{ax}), 0.95 (s, H₃-21), 1.61 (s, H₃, 23), 4.77 (q, J = 6.6 Hz, H-24), 1.34 (d, J = 6.6 Hz, H₃-26), 1.07 (d, J = 6.3 Hz, H-20), 0.85 (s, H₃-21), 0.85 (s, H₃-22), 1.16 (s, H₃, 23), 4.77 (q, J = 6.6 Hz, H-24), 1.34 (d, J = 6.6 Hz, H₃-4), ¹³C NMR (125 MHz, CDCl₃) δ 40.3 (C-1), 17.7 (C-2), 38.9 (C-3), 39.2 (C-4), 58.8 (C-5), 19.9 (C-6), 42.3 (C-7), 37.5 (C-8), 53.7 (C-9), 37.1 (C-10), 21.0 (C-11), 74.4 (C-12), 38.3 (C-13), 51.0 (C-14), 16.7 (C-15), 24.0 (C-16), 164.3 (C-17), 132.6 (C-18), 23.1 (C-19), 73.1 (C-20), 16.6 (C-21), 16.4 (C-22), 21.4 (C-23), 77.9 (C-24), 171.3 (C-25), 18.6 (C-26), 15.7 (C-27), 170.3 (CH₃CO), 21.8 (CH₃CO), 171.4 (C-1), 43.3 (C-2'), 64.2 (C-3'), and 22.2 (C-4').